X-Chromosomal Monosomy (77,X0) in a Doberman Pinscher With Gonadal Dysgenesis

F. W. K. Smith Jr, DVM, L. C. Buoen, BA, A. F. Weber, DVM, PhD, S. D. Johnston, DVM, PhD, J. F. Randolph, DVM, and D. J. Waters, DVM

A 6-month-old stunted female Doberman Pinscher was found to have a 77,X0 chromosomal complement. The ovaries were small, consisting primarily of interstitial-type cells and solid epithelial cords. The dam, sire, and a male littermate had normal karyotypes. (Journal of Veterinary Internal Medicine 1989; 3:90-95)

MONOSOMY X (X0) is a condition occurring in a conceptus following the union of egg and sperm, wherein one sex chromosome per cell is missing. It is usually due to a failure of either the male or female (spermatocyte or oocyte) chromosomes of a mating pair to separate at anaphase during meiosis. This results in the production of a phenotypic female with an X0 genotype.¹

Cases of X0 genotype have been reported in the human, Rhesus monkey, horse, sheep, pig, tammar wallaby, lesser bandicoot, Malayan black rat, mouse, and cat.²⁻¹⁴ Among Canidae, this condition has been reported only in the Silver Fox.¹⁵ Abnormalities observed in the various species vary considerably, and include those of the soma, and cardiovascular, urinary, and reproductive systems. Involvement of the genital tract is typically in the nature of small, afollicular ovaries, and hypoplastic uterine tubes, uterus, and vagina. In the bitch, the normal diploid chromosomal number is 76 autosomes plus two sex chromosomes (i.e., 78,XX). Only one of the two female (X) chromosomes is active in somatic cells and germ cells, except during oogenesis. The other X-chromosome becomes inactive and is recognized as sex chromatin (Barr body) in the nuclei of most somatic cells. In neutrophils, sex chromatin is recognized as a club-shaped nuclear appendage called a "drumstick."^{1.16}

The present report describes a case of gonadal dysgenesis in a 77,X0 dog, the first for this species.

Case Report

History

A 6-month-old female Doberman Pinscher weighing 16.4 kg was presented for ovariohysterectomy. A male littermate weighed 30 kg. Intermittent dysuria, poor weight gain, increased respiratory sounds that had improved with chloramphenicol administration, and chronic intermittent diarrhea were historic problems. Thoracic radiographs, which were obtained when the respiratory signs developed at 4 months of age, showed pleural fissure lines and increased peribronchial markings. Appetite and activity levels were normal. The dog was coprophagic, and had eaten linoleum. Vaccinations were current. The dog had never shown signs of estrus.

Physical Examination

The dog was of small stature, and was alert and responsive. Vital signs were normal. Physical abnormalities included increased bronchovesicular sounds noted on inspiration, thickened bowel loops, excessive skin in the ventrum of the neck,

From the Department of Clinical Sciences (Smith, Randolph), New York State College of Veterinary Medicine, Cornell University, Ithaca, New York; and the Departments of Veterinary Biology (Buoen, Weber) and Small Animal Clinical Sciences (Johnston, Waters), College of Veterinary Medicine, University of Minnesota, St. Paul, Minnesota.

Supported in part by funds from the Minnesota Agricultural Experiment Station (Project No. MIN-61-085).

Published as journal series No. 15427 as a contribution of the Minnesota Agricultural Experiment Station.

Reprint requests: Alvin F. Weber, DVM, PhD, University of Minnesota, Department of Veterinary Biology, College of Veterinary Medicine, 295 Animal Science/Veterinary Medicine Building, 1988 Fitch Avenue, St. Paul, MN 55108.

mucopurulent vaginal discharge, and an enlarged friable clitoris that protruded approximately 2 cm beyond the vulva. A semisoft stool was observed on rectal examination. There were no parasite eggs observed on a direct smear. There was one supernumerary nail located just dorsal to the normal nail on each pelvic limb digit.

Diagnostic Tests and Procedures

Hematologic and biochemical abnormalities included leukocytosis $(18.5 \times 10^3/\mu)$; normal, 6 to $17 \times 10^3/\mu)$, neutrophilia $(12.8 \times 10^3/\mu)$; normal, 3 to $11.5 \times 10^3/\mu)$, monocytosis $(1.7 \times 10^3/\mu)$; normal, 0.1 to $1.4 \times 10^3/\mu)$, hyperglycemia (114 mg/d); normal, 63 to 105 mg/d)), hypercalcemia (13.3 mg/d); normal, 5 to 12.9 mg/d)), and hyperphosphatemia (8.7 mg/d); normal, 4.3 to 7.6 mg/d)). These changes were considered stress or age related. The urine was concentrated (specific gravity, 1.046); the urinalysis results were unremarkable. An electrocardiogram was normal.

A pediatric vaginoscope could not be passed beyond the vestibule, suggesting agenesis or stricture of the vagina. Contrast vaginography identified an os clitoris, but failed to demonstrate a vagina.

At surgery, small ovaries $(0.5 \times 0.4 \times 0.5 \text{ cm} \text{ in size})$ were attached to the kidneys by connective tissue. There was one grossly normal uterine horn (4 mm in diameter) and a remnant left uterine horn. The os clitoris was excised in hope of resolving dysuria that might have been the result of distortion irritation of the vulva. The supernumerary nails were removed. Excessive intraoperative bleeding prompted evaluation for Von Willebrand's disease. Factor VIII-related antigen was less than 7% of the normal value. Blood was submitted for karyotyping, but was damaged during shipping. The dog was treated with ampicillin (20 mg/kg three times daily) for 10 days to prevent postoperative infection. Dysuria resolved postoperatively.

Histologically, the ovaries contained a well-vascularized cortex and medulla. The ovarian surface contained a mixed simple squamous to cuboidal epithelium, with frequent short epithelial cord and tubular invaginations typical of the dog ovary.¹⁷ Numerous long epithelial cords extended through the cortex into the medulla (Fig. 1A). A small aggregate of cuboidal cell-lined cavities resembling rete ovarii were present in the medulla. One of these was continuous with what appeared to be a medullary cord (Fig. 1B). No ovarian follicles or remnants of follicles or corpora lutea were observed. Ovarian interstitial cell masses were noted (Fig. 1C). Histologic studies made of ovaries from three approximately 6-month-old dogs (Doberman Pinscher \times Collie; Husky; and Schnauzer) showed many primordial and larger follicles (Figs. 2A and 2B) and occasional degenerate tertiary (Graafian) follicles (Fig. 2C). Minimal uterine glandular development (Fig. 1D), similar to that observed previously in an X0 cat, was found in the normalappearing uterine horn.14

Serum collected before ovarihysterectomy was submitted for testosterone, estradiol, progesterone, luteinizing hormone (LH), and thyroxine (T₄) determinations. The testosterone concentration was elevated at 1.02 ng/ml (normal, <0.4 ng/ml). The estradiol concentration was 47.80 pg/ml, the progesterone concentration was 0.28 ng/ml, and the LH concentration was 2.5 ng/ml. The T₄ concentration was elevated at $3.52 \ \mu$ g/dl (normal, 1.5 to 3.0 μ g/dl). The dog was presented 2 months later for an intestinal obstruction, which was related to ingestion of parts of a couch. This obstruction was corrected through multiple enterotomies, without excessive bleeding. A repeated Von Willebrand's test showed 23% of normal levels of Factor VIII-related antigen.

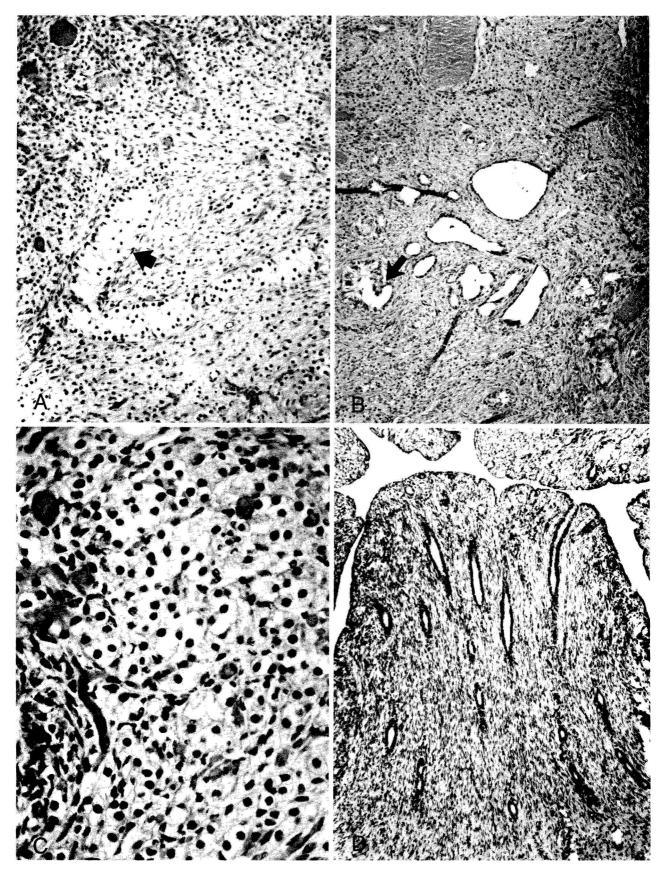
Heparinized blood and a skin biopsy specimen were collected for cytogenetic and neutrophil drumstick evaluations using techniques described previously.¹⁸ A karyotype of 77,X0 (monosomy-X) was found in both the blood and skin (Fig. 3), while the dam, sire, and male littermate had normal karyotypes. A normal bitch should have a 78,XX karyotype with approximately 4% drumsticks.¹⁶ Neutrophil drumsticks (inactivated X-chromosome) were not observed in the dog.

Discussion

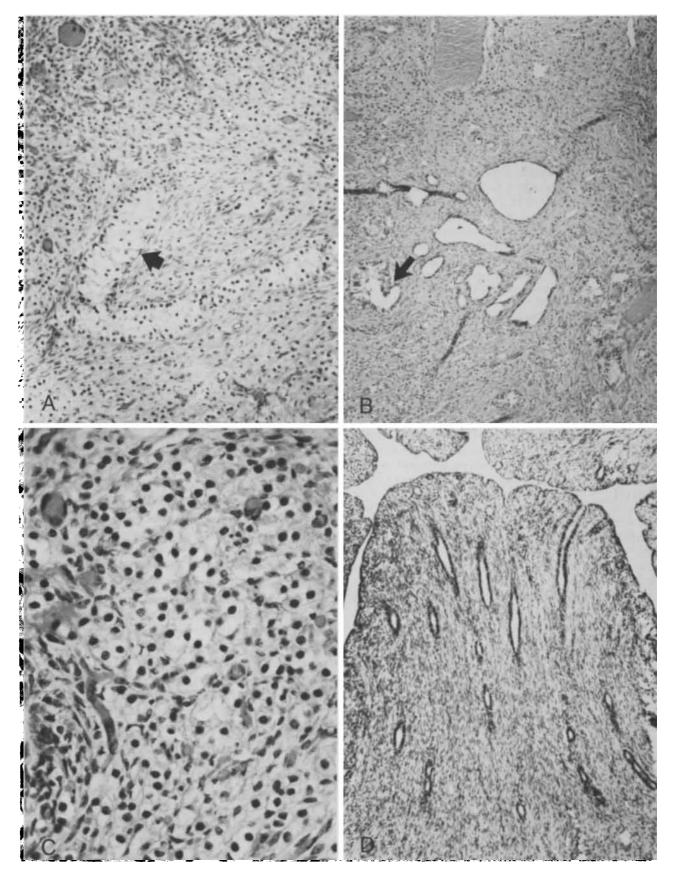
Many of the morphologic abnormalities in this dog can be attributed to the X0 genotype. Small stature and hypoplasia of the genital system is commonly reported in animals of other species with monosomy-X. Metabolic causes for stunted growth were not supported by our laboratory test results. Webbing of the neck also is commonly observed in women with the X0 chromosome complement.¹⁹ Supernumerary nails have not been reported in other cases of monosomy-X. However, women with an X0 chromosome complement may have hypoplastic nails, and 50% have short fourth metacarpals.¹⁹ The presence of an os clitoris has not been described previously with this condition and would suggest elevated testosterone concentrations during development. This is supported by the increased serum testosterone values observed. The source of testosterone is not apparent.

The dysuria was believed to be due to the enlarged clitoris, causing distortion and irritation of the vulva. The dysuria resolved following partial amputation of the clitoris and removal of the os clitoris. Von Willebrand's disease, a condition common in the Doberman Pinscher,²⁰ probably accounted for the excessive bleeding during the first surgical procedure. The increase in the percentage of Factor VIII-related antigen on the second test may reflect either laboratory variation, a true age-related increase,²⁰ or patient variability.

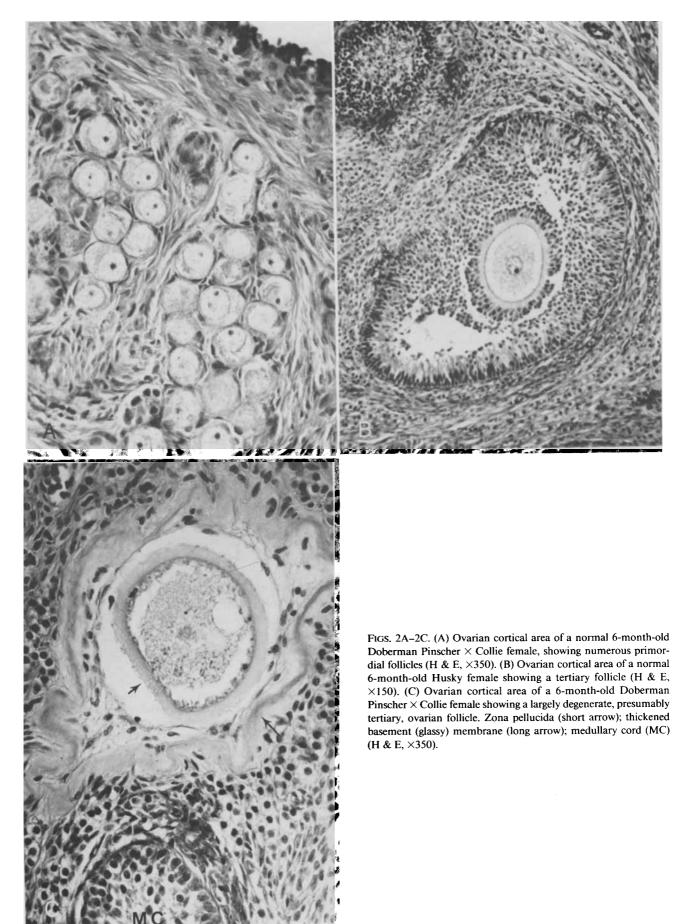
The elevated T_4 concentration in this dog is probably a normal variant. The estradiol concentration suggested proestrus. This was not supported by histopathologic evaluation of the ovaries. The dog was not exhibiting proestrus behavior and there was no genital tract bleeding. Cytologic evaluation of a smear from the vestibule revealed no abnormalities. Possible causes for these near-normal findings would include ectopic functional gonadal tissue, unobserved follicles in either ovary, or laboratory error. The progesterone concentration was compatible with anestrus, proestrus, or estrus. The LH concentration could reflect normal endogenous pulsatile activity in an anestrus bitch, or a high baseline level due to gonadal insufficiency.

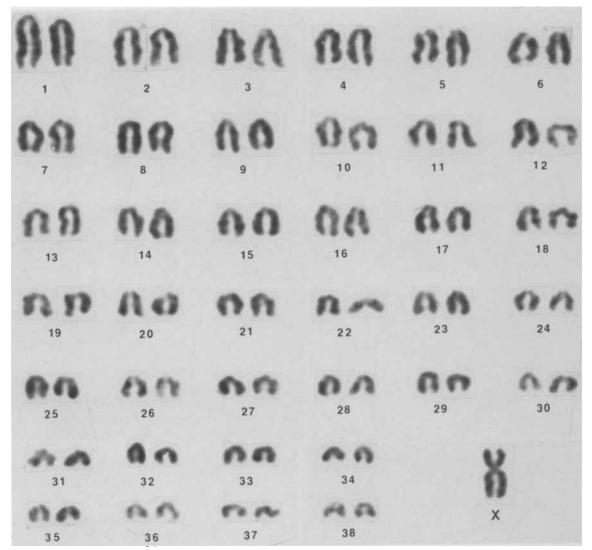


FIGS. 1A-1D. (A) Ovarian cortex of a Doberman Pinscher with X-chromosomal monosomy, showing solid epithelial cords (H & E, $\times 150$). (B) Ovarian medulla of a Doberman Pinscher with X-chromosomal monosomy. Arrow shows continuity between apparent medullary cord and rete ovarii structures (H & E, $\times 100$). (C) Ovarian cortex of a Doberman Pinscher with X-chromosomal monosomy, showing ovarian interstitial cell masses (H & E, $\times 350$). (D) Endometrium of a Doberman Pinscher with X-chromosomal monosomy. Note low uterine surface epithelium and minimal uterine glandular development (H & E, $\times 150$).



FIGS. 1A-1D. (A) Ovarian cortex of a Doberman Pinscher with X-chromosomal monosomy, showing solid epithelial cords (H & E, $\times 150$). (B) Ovarian medulla of a Doberman Pinscher with X-chromosomal monosomy. Arrow shows continuity between apparent medullary cord and rete ovarii structures (H & E, $\times 100$). (C) Ovarian cortex of a Doberman Pinscher with X-chromosomal monosomy, showing ovarian interstitial cell masses (H & E, $\times 350$). (D) Endometrium of a Doberman Pinscher with X-chromosomal monosomy. Note low uterine surface epithelium and minimal uterine glandular development (H & E, $\times 150$).





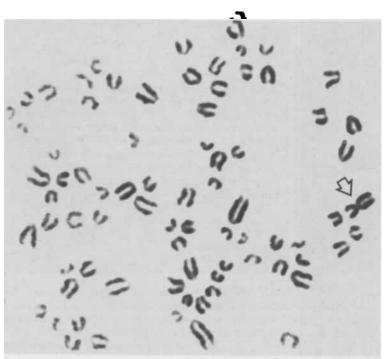


FIG. 3. Karyotype (above) and chromosome spread (below) of a Doberman Pinscher with X-chromosomal monosomy (77,X0). Arrow in chromosome spread shows X-chromosome (Giemsa stain). A point of interest in this report of monosomy-X (77,X0) in a 6-month-old dog is whether or not follicular development occurred postnatally. The presence of afollicular cortical epithelial cords and surface epithelial invaginations along with medullary cord and rete-like structures suggests that ovarian follicular development may not have continued normally (Figs. 2A-2C), but instead, early postnatal dysgenesis ensued.

One of the initial explanations for the pathogenesis of gonadal dysgenesis was that low X-gene dosage has a deleterious effect on oocvtes. Therefore, follicular formation occurs early, but follicular atresia develops gradually postnatally.²¹ Consequently, X0 rodents with a shorter reproductive span would be fertile initially, while in larger species, such as humans, sterility would occur long before puberty.²² However, the reports of occasional fertile X0-women and sterility in the X0-cat contradict this theory.^{14,23} Miklos originally,²⁴ and Burgoyne and Baker,²⁵ and Speed²⁶ more recently, suggested that meiotic cells are selectively spared by some mechanism when homologous chromosomes are acrocentric in type. This is explained on the basis that in such a chromosome, nonhomologous ("fold-back") pairing of sister chromatids onto themselves may occur resulting in the X-chromosome forming a "hairpin" configuration. Thus, X0-oocytes from species with acrocentric X-chromosomes (e.g., mouse) would be spared. In the human, horse, dog, fox, and cat, which have metacentric X-chromosomes, fold-back pairing would be less able to occur. Thus, oocyte atresia would occur earlier in these species. If atresia occurred sufficiently early in the case reported here, evidence of a prior existence of follicular structures, as observed in normal development (Fig. 2C), would be lost. The epithelial aggregates of the second proliferation, which would form the definitive follicles, would remain as afollicular cords rather than subdividing into follicles. The presence of such afollicular cords, plus large aggregates of interstitial cells of the ovary, is what characterized the ovarian sections of the Doberman Pinscher in this report.

References

- Grumbach MM, Van Wyk JJ. Sex chromosome anomalies and disorders of gonadal differentiation. In: Williams RH, ed. Textbook of Endocrinology. Philadelphia: WB Saunders, 1981; 423-514.
- 2. Turner HH. A syndrome of infantilism, congenital webbed neck, and cubitus valgus. Endocrinology 1938; 23:566-574.

- Weiss G, Weick RF, Knobil E, et al. An X0 anomaly and ovarian dysgenesis in a Rhesus monkey. Folia Primatol (Basel) 1973; 19:24-27.
- Chandley AC, Fletcher J, Rossdale PD, et al. Chromosome abnormalities as a cause of infertility in mares. J Reprod Fertil 1975; (Suppl) 23:337–383.
- 5. Hughes JP, Benirschke K, Kennedy PC, et al. Gonadal dysgenesis in the mare. J Reprod Fertil 1975; (Suppl) 23:385-390.
- Zartman DL, Hinesley LL, Gnalkowski MW. A 53,X female sheep (*Ovis aries*). Cytogenet Cell Genet 1981; 30:54–58.
- Nes N. Betydningen av kromosomaberrasjoner hos dyr. Forskn Fors Landbruket 1968; 19:393-410.
- Sharman GB, Robinson ES, Walton SM, et al. Sex chromosomes and reproductive anatomy of some intersexual marsupials. J Reprod Fertil 1970; 21:57-68.
- Sharma T, Raman R. An X0 female in the Indian mole rat. J Hered 1971; 62:384–387.
- Yong H. Presumptive X monosomy in black rats from Malaya. Nature 1971; 232:484–485.
- Russel WL, Russel LB, Gower JS. Exceptional inheritance of a sex-linked gene in the mouse explained on the basis that the X/0 sex-chromosome constitution is female. Proc Natl Acad Sci USA 1959; 45:554-560.
- Nordby DE, Hegreberg GA, Thuline HC, et al. An X0 cat. Cytogenet Cell Genet 1974; 13:448–453.
- Long SE, Berepubo NA. A 37,X0 chromosome complement in a kitten. J Small Anim Pract 1980; 21:627-631.
- Johnston SD, Buoen LC, Madl JE, et al. X-chromosomal monosomy (37,X0) in a Burmese cat with gonadal dysgenesis. J Am Vet Med Assoc 1983; 182:986–989.
- Makinen A, Valtonen M. The X0 syndrome in the silver fox (Vulpes fulvus Desm.) Presented at the Sixth European Colloquim on Cytogenetics of Domestic Animals, Zurich, Switzerland, 1984.
- Colby EB, Calhoun L. Accessory lobule on the polymorphonuclear neutrophil leukocyte of domestic animals. Acta Cytol 1963; 7:346-350.
- Ellenberger W. Handbuch der Vergleichenden Mikroskopischen Anatomie der Haustiere. Zweiter Band. Berlin: Verlagsbuchhandlung Paul Parey, 1911; 662.
- Lin CC, Newton DR, Smink WK, et al. A rapid and simple method for the isolation and culture of leukocytes for chromosome analysis in domestic animals. Can J Anim Sci 1976; 56:27-31.
- Wilson JD, Griffin JE III. Disorders of sexual differentiation. In: Braunwald E, et al. ed. Harrison's Principles of Internal Medicine. New York: McGraw Hill, 1987; 1840–1853.
- Green RA. Bleeding disorders. In: Ettinger SJ, ed. Textbook of Veterinary Internal Medicine. Diseases of the Dog and Cat. Philadelphia: WB Saunders, 1983; 2076–2098.
- Fayez JA. The pure gonadal dysgenesis syndrome. Int J Gynaecol Obstet 1978; 15:550-553.
- Burgoyne PS. The role of sex chromosomes in mammalian germ cell differentiation. Ann Biol Anim Biochem Biophys 1978; 18:317-325.
- 23. Nakashima I, Robinson A. Fertility in a 45,X female. Pediatrics 1971; 47:770-773.
- Miklos GLG. Sex-chromosome pairing and male fertility. Cell Genet 1973; 13:558-577.
- Burgoyne PS, Baker TG. Perinatal oocyte loss in X0 mice and its implications for the aetiology of gonadal dysgenesis in X0 women. J Reprod Fert 1985; 75:633-645.
- Speed RM. Oocyte development in X0 foetuses of man and mouse: The possible role of heterologous X-chromosome pairing in germ cell survival. Chromesoma 1986; 94:115-124.